Transdermal Absorption of Castor Oil

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Abstract

With the current high level of interest in patches to deliver drugs across the skin, the concept of external applications for delivery of medicinal substances is becoming widely accepted. This study evaluated the potential for transdermal absorption of castor oil, a treatment modality recommended by Edgar Cayce. Specific epoxydicarboxylic acids are known to be excreted via the urine when castor oil is administered orally. Comparison of the amounts of these substances in the urine of subjects given oral and transdermal administrations of castor oil may provide evidence of transdermal absorption of castor oil and provide clues to its metabolism in the system.

Three volunteers were given castor oil via abdominal hot pack and oral administration. Urine samples were collected before and after the interventions. Oral administration of castor oil resulted in high levels of excretion of the epoxydicarboxylic acids in all the subjects, results similar to a previous study. In contrast, the level of urinary epoxydicarboxylic acids in the external application (abdominal pack) sessions did not vary greatly from the relatively low endogenous levels of these molecules that are normally present. If the castor oil was absorbed through the skin and metabolized in the system, it did not have the effect of significantly increasing the excretion of the specific metabolic byproducts associated with ingestion of castor oil. The implications of these findings are discussed and directions for future research are considered.

Introduction

Castor oil has a long history as a medicinal agent in folk healing and mainstream medicine. Its primary function has been as an oral laxative.

In his book *The Oil That Heals* (1993), McGarey detailed numerous case reports supporting the therapeutic efficacy of externally applied castor oil packs for a wide range of disorders. McGary's review relies heavily on the Edgar Cayce legacy. The work of Edgar Cayce, an important figure in the development of holistic medicine in the modern era (Callan, 1979; Mein, 1989), places great significance on healing properties of castor oil applied externally as a pack. The typical application recommended by Cayce is that of a hot abdominal pack placed over the right side of the abdomen covering the liver, upper small intestines, cecum, and ascending colon.

One of the major drawbacks for using the Cayce regimen for castor oil packs is that they tend to be messy and difficult to apply. With the current high level of interest in patches to deliver drugs across the skin, the concept of external applications for delivery of medicinal substances is becoming widely accepted. The intent of this study was to develop a reliable means for measuring levels of transdermal absorption of externally

applied castor oil. Identifying one or more chemical markers could then allow for comparison of Cayce's typical application with more modern systems of dermal delivery that tend to be less messy and more practical.

Although researchers have developed techniques for measuring the amount of castor oil absorbed into the system when it is taken orally, we have found no studies that have quantified the absorption levels for externally applied castor oil. Thus, we believe our study is the first attempt to measure the absorption of castor oil administered through the skin.

A previous study done by Hagenfeldt et al. (1986) provided a conceptual framework for our research design. In the Hagenfeldt et al. study, castor oil was administered orally to three healthy volunteers. Urine was collected for several hours and analyzed using capillary gas chromatography. The analysis revealed elevated levels of the products of breakdown of castor oil by the liver (specific epoxydicarboxylic acids that are normally found in urine in very small amounts). Thus, the presence of high levels of these acids is evidence for absorption and metabolism of orally administered castor oil.

Using the data and conceptual model from the Hagenfeldt et al. study, we designed a study that included both oral and external (abdominal pack) administration of castor oil. By comparing the levels of the epoxydicarboxylic acids in the urine following external and oral applications, we hoped to be able to reliably quantify the level of absorption of the oil through the skin.

Methods

Design

This preliminary study utilized a within-subjects design with pre and post intervention data collection.

Subjects

Three healthy Meridian Institute researchers volunteered for this project. All participants had used abdominal castor oil packs previously and knew how to do the procedure. None of the participants had used castor oil packs for at least several months before the study.

Procedures

In the first stage of the project, urine samples were collected before and during two three-day series of abdominal castor oil packs. After a one-week washout period in which urine samples were collected to determine clearance rate, castor oil was administered orally on two occasions (one week apart) with urine samples collected at baseline and after the ingestion of castor oil. For the first oral administration of castor oil, 2.5 cc was given. For the second oral administration was 15 cc was given.

The procedure for the abdominal castor oil pack sessions is documented in Appendix A. Since the level of heat used in the abdominal castor oil pack may be a factor in transdermal absorption, three levels of heat were applied using electrical heating pads (50 watts, 100 watts, and 200 watts). The following sequence of heating was used in the first stage of the project:

	First Series	Second Series
Subject 1	No heat	100 watt
Subject 2	50 watt	200 watt
Subject 3	50 watt	50 watt

All urine samples were tested for the presence and concentration of two epoxydicarboxylic acids (epoxyoctanedioic acid and epoxydodecanedioic acid) and ricineolic acid. The urine samples were analyzed by Alturas Analytics, Inc. (Moscow, ID) using the capillary gas chromatography as described by Hagenfield et al (1986). The sequence of interventions and sampling for Stage 1 is provided in Table 1 (Appendix B).

The second stage of the project was essentially a focused and scaled down version of the first stage to confirm the findings and decrease sources of error. A different laboratory (EnviroTest Laboratories, Edmonton, AB, Canada) was used to minimize laboratory practices as a source of variability. The highest level of heating pad was used for all sessions to facilitate maximum penetration of the castor oil into the skin. Dietary oils of a similar composition to castor oil were eliminated (e.g., olive oil) at least one week prior to and during the second stage. Since no detectable levels of ricineolic acid was found in any of the samples in stage 1, this substance was not included in the lab assays for stage 2. The sequence of interventions and sampling for Stage 2 is provided in Table 2 (Appendix B).

Results

For the oral administration of castor oil in either of the two amounts, there was a high level of excretion of the epoxydicarboxylic acids in all the subjects, results similar to the Hagenfeldt et al. study.

In contrast, the level of urinary epoxydicarboxylic acids in the external application (abdominal pack) sessions did not vary greatly from the relatively low levels of these molecules normally present. The amount of heat used did not affect the outcome.

Discussion

Considering how permeable the skin is to a wide range of substances, it would be truly amazing if no significant amounts of castor oil were absorbed after an hour and a half of contact with high heat. Yet we were unable to show this based on measuring the products produced in the liver and excreted in the urine, which are very evident when castor oil is taken orally.

We took care to increase the chances of transdermal absorption in Stage 2 by increasing the level of heat to the maximum (200 watt heating pads), prolonging all castor oil sessions to one and one-half hours in duration, and avoiding possible confounding dietary sources of oils similar to castor oil (especial olive oil). Apparently these adjustments did not have a notable affect on the outcome that is essentially in agreement with the Stage 1 results.

The Cayce readings do consistently maintain that small amounts of substances frequently have greater therapeutic effects than large doses. This concept is similar to the homeopathic view on the relative potency of dosages wherein small amounts of chemicals can have a greater or different effect than large amounts. Perhaps a small amount of castor oil absorbed through the skin has a different effect than the same amount absorbed through the gut. When taken orally, castor oil is broken down with powerful enzymes in the small intestine before being sent to the liver for further processing. The raw castor oil could have a local effect directly on the underlying tissues before cycling through the liver for processing, which may take longer without the benefit of intestinal enzymes. Thus, low levels of castor oil could have a therapeutic effect without producing a spike in urinary epoxydicarboxylic acids.

To help clarify our findings and give us clues as to how to proceed with the next phase of investigation, we contacted the International Castor Oil Association and several of its members requesting data or insights into the possible transdermal properties of castor oil. So far we have not received any useful suggestions for the further exploration of how castor oil is absorbed through the skin and metabolized in the system.

One possibility for further inquiry is to see if ricinoleic acid, the major component of castor oil, is found in the blood immediately after the application of a castor oil pack. If it is not, then apparently the castor oil does not cross the skin barrier. If it is found in the blood, but the breakdown products are not found in the urine in detectable amounts (as we have already determined), then it is metabolized in a way not yet understood. This could offer significant opportunities for further research on the biochemistry and metabolism of castor oil and its relation to healing.

Another possibility noted by McGarey is that castor oil is assimilated into the system *vibratorially*. Pondering the idea that everything is composed of constantly moving subatomic particles, McGarey asserts that, "All substances, then, whether they be living or not, give off vibrations ... Is it really vibration, then, that carries the healing nature of castor oil into the body ...?" (McGarey, 1993, p. 30). If this is the case, the question naturally arises as to how one measures the vibratory effects of a castor oil pack.

References

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Appendix A: Instructions For Castor Oil Packs

To Make The Pack

Take the piece of wool flannel and fold it into 3 thicknesses. Put it in a pan (like a large disposable baking pan) and pour castor oil on it. Saturate the whole flannel, and leave it until it is well-saturated. When you use it, you want it saturated, but not dripping. The size you want for use is about 12 x 18 inches - about the same size as your heating pad, so that the pad heats the whole pack, but does not touch your skin. After each use, you will probably need to add a little more castor oil.

Where To Place The Pack

Place the pack on the right side of the body, extending from a little above the bottom of the sternum (breastbone) to about 4 inches below the navel. It should go from the navel around the body on the right side as far to the side as you can get it. Basically, cover as much of your right side as you can.

Using The Pack

Use the pack in the evening, as you are resting before bed. Spread out a large plastic garbage bag on the bed so that the castor oil won't leak onto the bed. Fold a towel (that you will use only for castor oil packs, because the oil is almost impossible to wash out completely). Take the cloth cover off the heating pad. Then put the heating pad on top of the towel.

Heat the pack before you put it on yourself. One way is to heat it in the oven on a "low" temperature setting. Another is to heat it in a microwave oven (in a microwave-safe container) for about one minute. Another way is to put the pack on top of the heating pad and turn the pad on high. Let it warm up for a few minutes.

Lie down on your back on the plastic bag. Place the pack on your abdomen, with the heating pad on top and the towel on top of that. Have the heating pad control within easy reach of your hand, because you may need to adjust it so the pack is not too hot or too cold. It should be very warm, but not so hot it burns you. Be careful! If you're not sure how hot is safe, start with a slightly warm session and work up to a warmer one. If it's not warm enough, it may help to rest your arms and hands on the towel to press the pack onto your body.

Keep the pack on for 1 to 1½ hours. Have a paper towel handy to wipe the oil off yourself when you get up, and be careful not to get any on the bed. Make up a solution of baking soda in warm water (2 teaspoons to a quart), and use paper towels or a sponge with the baking soda solution to clean off the castor oil. You might also want to take a shower with soap after that.

Use the pack for 3 days in a row.

After 3 Days Of Using The Pack

On the evening of the 3rd day, take olive oil (not castor oil!) by mouth. Begin with about a teaspoon. If you feel OK, you can take up two tablespoons.

Appendix B: Tables

Table 1 (Stage 1)

		Subject 1				Subject 2			Subject 3		
	Activity	Samples	E1	E2	R	E1	E2	R	E1	E2	R
Day 1		Sample 1	234300	290400	ND	341250	339000	ND	629000	783000	ND
Day 2	CO Pack 1										
Day 3	CO Pack 2										
Day 4	CO Pack 3	Sample 2	253500	240500	ND	244200	156600	ND	1616000	2144000	ND
Day 5											
Day 6											
Day 7											
Day 8											
Day 9	CO Pack 4	Sample 3	198000	153000	ND	760000	127600	ND	588500	621500	ND
Day 10	CO Pack 5										
Day 11	CO Pack 6	Sample 4	507600	712800	ND	527400	221400	ND	365400	384300	ND
Day 12		Sample 5 (S1)	596750	470250	ND						
Day 13		Sample 5 (S2)				497700	274500	ND	515100	709750	
Day 14		Sample 5 (S3)									ND
Days 15-22	Washout										
Day 23		Sample 6	604800	803200	ND	177600	261600	ND	309350	353050	ND
Day 24	2.5cc CO	Sample 7	2915200	30240000	ND	11610000	31050000	ND	14522500	50505000	ND
Days 25-30	Washout										
Day 31	15cc CO	Sample 8	4972500	103350000	ND	7328000	32640000	ND	38700000	86940000	ND

Note: All results expressed as total acids (ng/ml x urine volume) E1 = Epoxyoctanedioic Acid E2 = Epoxydodecanedioic Acid

R = Ricineolic Acid (not detected)

Table 2 (Stage 2)

			Subject 1		Subj	ject 2	Subject 3	
	Activity	Samples	E1	E2	E1	E2	E1	E2
Day 1		Sample 1	2415	360	4078.8	183.6	5592.65	527.8
Day 2	CO Pack 1	Sample 2	1203.5	130.5	3150	199.5	1759.5	218.5
Day 3	CO Pack 2							
Day 4	CO Pack 3	Sample 3	4704	441	2621.5	192.5	2464.15	211.85
Day 5		Sample 4	1863	207	8257.4	590.4	4033.8	547.8
Days 6-13	Washout							
Day 14		Sample 5	2329	306	1956	96	5886	634.5
Day 15	2.5cc CO	Sample 6	75313	44602	40802.5	12820.3	84700	45850